

Year	1970	1971	1972	1973	1974	1975	1976	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	2052	2053	2054	2055	2056	2057	2058	2059	2060	2061	2062	2063	2064	2065	2066	2067	2068	2069	2070	2071	2072	2073	2074	2075	2076	2077	2078	2079	2080	2081	2082	2083	2084	2085	2086	2087	2088	2089	2090	2091	2092	2093	2094	2095	2096	2097	2098	2099	2100
1970	1971	1972	1973	1974	1975	1976	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	2052	2053	2054	2055	2056	2057	2058	2059	2060	2061	2062	2063	2064	2065	2066	2067	2068	2069	2070	2071	2072	2073	2074	2075	2076	2077	2078	2079	2080	2081	2082	2083	2084	2085	2086	2087	2088	2089	2090	2091	2092	2093	2094	2095	2096	2097	2098	2099	2100	

Respectfully submitted,

Yongzhi Yang
Registration No. (see attached)

**INTELLECTUAL PROPERTY/
TECHNOLOGY LAW**
P. O. Box 14329
Research Triangle Park, NC 27709
(919) 419-9350
Attorney Reference: 4176-101

APPENDIX A

Version with Markings to Show Changes Made

In the Specification:

1. On page 2, the paragraph beginning at line 12 has been changed, as follows:

ELPs, as explained more fully in the Detailed Description of the Invention hereof (Section 5) are oligomeric repeats of the pentapeptide Val-Pro-Gly-X-Gly (Sequence ID No. 1), where the guest residue X is any amino acid. ELPs undergo a reversible inverse temperature transition. They are highly soluble in water below the inverse transition temperature (T_i), but undergo a sharp (2-3°C range) phase transition when the temperature is raised above their T_i , leading to desolvation and aggregation of the polypeptide.^{1, 2, 3} In previous work, McPherson et al. have exploited the inverse transition to purify recombinant poly(GVGVP) polypeptides. Previous studies have also shown that protein conjugates of poly(N-isopropylacrylamide), a synthetic polymer that undergoes a similar thermally-reversible phase transition, also retain the transition behavior of the free polymer.^{5, 6, 7}

2. On page 14, the paragraph beginning at line 15 has been changed, as follows:

Another preferred ELP comprises polymeric units having the sequence IPGXG (Sequence ID No. 2), where X is as defined above.

3. On page 23, the paragraph beginning at line 26 has been changed, as follows:

The objective in this example was to design a β -turn sequence with a predicted T_i above 37°C so that an FP would remain soluble under conditions used for E. coli culture, but which could be aggregated by a small increase in temperature. Previous studies by Urry and colleagues have shown that two ELP-specific variables, guest residue(s) composition²⁸ (i.e., identity and mole fraction of X in the VPGXG monomer) and chain length²⁹ of the ELP profoundly affect the transition temperature, and thereby provide design criteria to specify the T_i for a specific application. Based on these studies, a gene was synthesized encoding an ELP sequence (Sequence ID No. 3) with guest residues valine, alanine, and glycine in the ratio 5:2:3, with a predicted T_i of ~40°C in water. The synthetic gene, which encoded 10 VPGXG pentapeptide repeats (the "10-

mer”), was oligomerized up to 18 times to create a library of genes encoding ELPs with precisely-specified molecular weights (MWs) ranging from 3.9 to 70.5 kDa. To my knowledge, these are the first examples of genetically-engineered ELPs with precisely-defined chain length and amino acid sequence, which are designed to exhibit an inverse transition at a specified temperature. Thioredoxin was expressed as a N-terminal fusion with the 10-, 20-, 30-, 60-, 90-, 120-, 150-, and 180-mer ELP sequences, and tendamistat was expressed as a C-terminal fusion to thioredoxin/90-mer ELP (Figure 1b).

4. On page 32, the paragraph beginning at line 21 has been changed, as follows:

Standard molecular biology protocols were used for synthesis and oligomerization of the ELP genes (Ausubel, et al.³²). Monomer genes for two ELP sequences were utilized in this example. The first, ELP[V₅A₂G₃-10] encoding ten Val-Pro-Gly-Xaa-Gly repeats where Xaa was Val, Ala, and Gly in a 5:2:3 ratio, respectively, had been synthesized previously³⁷. The second monomer, ELP[V-5] (Sequence ID No. 4), encoded five Val-Pro-Gly-Val-Gly pentapeptides (i.e., Xaa was exclusively Val). The coding sequence for the ELP[V-5] monomer gene was: 5'-GTGGGTGTTCCGGGCGTAGGTGTCCCAGGTGTGGGCGTACCGGGCGTTGGTGTTCCTG GTGTCGGCGTGCCGGGC-3' (Sequence ID No. 5). The monomer genes were assembled from chemically synthesized, 5'-phosphorylated oligonucleotides (Integrated DNA Technologies, Coralville, IA), and ligated into a pUC19-based cloning vector. A detailed description of the monomer gene synthesis is presented elsewhere³⁸.

5. On page 33, the paragraph beginning at line 11 has been changed, as follows:

Different ELP constructs are distinguished here using the notation ELP[X_iY_j-n], where the bracketed capital letters are single letter amino acid codes and their corresponding subscripts designate the frequency of each guest residue in the repeat unit, and n describes the total length of the ELP in number of pentapeptides. The two ELP constructs central to the present example are ELP[V₅A₂G₃-90] (35.9 kDa) (Sequence ID No. 6) and ELP[V-20] (9.0 kDa) (Sequence ID No. 7).